## Abstract

Inositol monophosphatase (IMPase) is a  $Mg^{2+}$  activated and  $Li^+$  inhibited phosphatase that hydrolyzes phosphomonoester bond of *myo*-inositol-1-phosphate (I-1-P). IMPase plays a crucial role in the development and proper functioning of the brain due to its involvement in phosphoinositol signalling cascade. IMPase is an important target of  $Li^+$ -based therapeutics for manic depressive disorders. Despite the therapeutic efficiency of  $Li^+$ , the precise mechanism of the  $Li^+$ -induced inhibition of IMPase remains obscured. IMPase shows the highest catalytic activity with the substrate I-1-P, but it also manifests substrate promiscuity with other substrates like 2'AMP and  $\beta$ -glycerophosphate. Attempts have been made with staphylococcal IMPase-I (SaIMPase-I; SAS2203) and IMPase-II (SaIMPase-II; SAS1042) to decipher the (i) molecular basis of substrate promiscuity, (ii) mode of inhibition by metal ions like  $Li^+$  and high concentration of Mg<sup>2+</sup> and (iii) possible role of SaIMPase-II in staphylococcal biofilm formation.

Crystal structures of SaIMPase-II and its substrate (I-1-P and 2'AMP) bound complexes have revealed the role of active site mobile loops (connecting  $\alpha 1-\alpha 2$ ,  $\alpha 4-\beta 6$ ,  $\alpha 6-\beta 8$  and  $\alpha 5-\beta 7$ ) in substrate selection. Furthermore, considering the high degree of structural homology and conserved active site residues of SaIMpase-I with human IMPase-I, SaIMPase-I is used as a model enzyme to study molecular details of Li<sup>+</sup> inhibition. Li<sup>+</sup> inhibition in IMPase is competitive with Mg<sup>2+</sup> and shares a common or overlapping binding site. Li<sup>+</sup> is virtually invisible in X-ray crystallography since Li<sup>+</sup> has only two electrons to interact with X-ray. The crystal structure of SaIMPase-I ternary product complex (SaIMPase-I/3Mg<sup>2+</sup>/PO<sub>4</sub><sup>3-</sup>) shows a bound phosphate and three Mg<sup>2+</sup> (namely Mg1, Mg2 and Mg3) in the active site. The competitive displacement of Mg<sup>2+</sup> as a function of an increasing LiCl concentration in the crystals of SaIMPase-I ternary product complex has been employed to visualize the binding site of Li<sup>+</sup>. The complete absence of the electron density corresponding to the pre-occupied Mg2 is observed in LiCl soaked structures. Moreover, based on a detailed investigation of the phosphate conformation and the coordination state of the bound  $Mg^{2+}$  ions, the inhibition mechanisms of  $Li^+$  and high concentrations of Mg<sup>2+</sup> are proposed. It is also reported that the SaIMPase-II is essential in staphylococcal polysaccharide-independent biofilm formation. Overexpression of SaIMPase-II in E. coli produces extracellular macroscopic fibers and it is confirmed that the fibers are composed of SaIMPase-II. The amyloidic nature of the fiber has been evaluated by high-resolution electron microscopy complemented with X-ray fiber diffraction and binding of amyloid-specific dyes, such as Congo red and Thioflavin-T. The fibers are found to be highly sticky in nature and bind a large number of bacterial cells. It is suggested that SaIMPase-II is involved in fibers formation and acts as the adhesion in biofilm matrix.

## Key words

Inositol monophosphatase; *Staphylococcus aureus*; crystal structure; substrate specificity; Li<sup>+</sup> inhibition; amyloid fiber.